

# NUOVI FARMACI E TRAPIANTO

Udine 21-22 gennaio 2016

## Integrazione dei nuovi farmaci nel programma trapiantologico della leucemia linfoblastica acuta: UN CASO CLINICO...PEDIATRICO

### “Which ALL” is eligible for HSCT?

- 10% of children with very high risk ALL in CR1
  - all S3/S4 CR2 (early medullary relapses)
  - S2 CR2 with high MRD after induction
- are eligible for allogeneic stem cell transplantation

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## **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

### **4 standard approaches**

> 1985

Palliative care

Chemotherapy

DLI

Second transplant

## **PEDIATRIC ALL “MOLECULAR RELAPSING” AFTER TRANSPLANT**

## **PEDIATRIC ALL HIGH MOLECULAR MRD LEVEL BEFORE TRANSPLANT**

# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

*Relapse after HCT in pediatric ALL in CR1 & CR2 EBMT Results – Myriam Labopin*

**3628 pediatric ALL** in CR1 (45%) & CR2 (55%) reported to the EBMT in 10ys

**23%** out of 3628 relapse at a median of **6.5 ms** (range 1-67; 25<sup>th</sup>: 4; 75<sup>th</sup>: 12 ms)

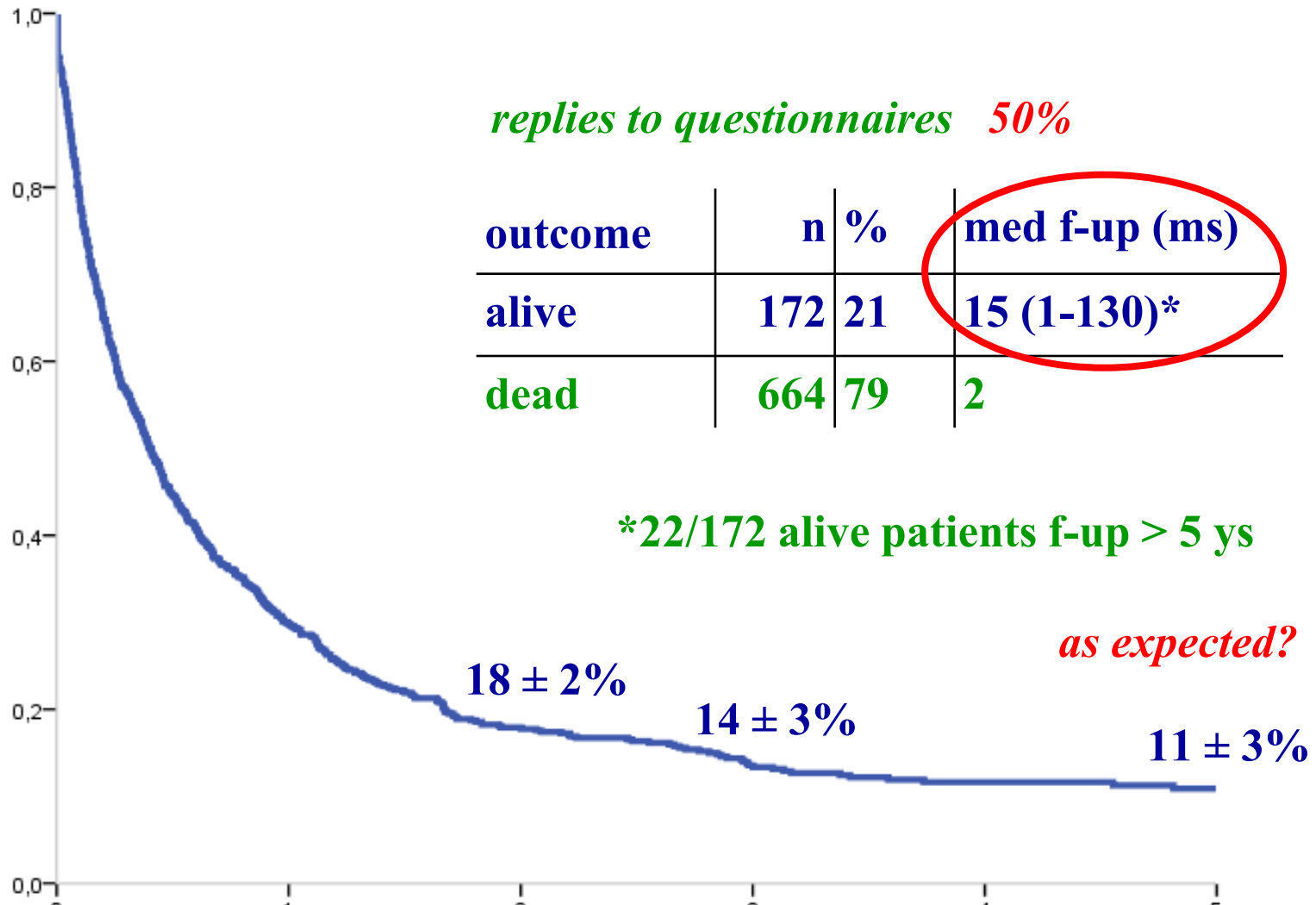
**2 yr cumulative incidence of relapse after HCT** **25%** (SE 1)

incidence of relapse in CR1 **21%** and CR2 **26%**

enrolment **n = 836** (M 66%, median 9 ys)

# Relapse after HCT in pediatric ALL in CR1 & CR2 - Results

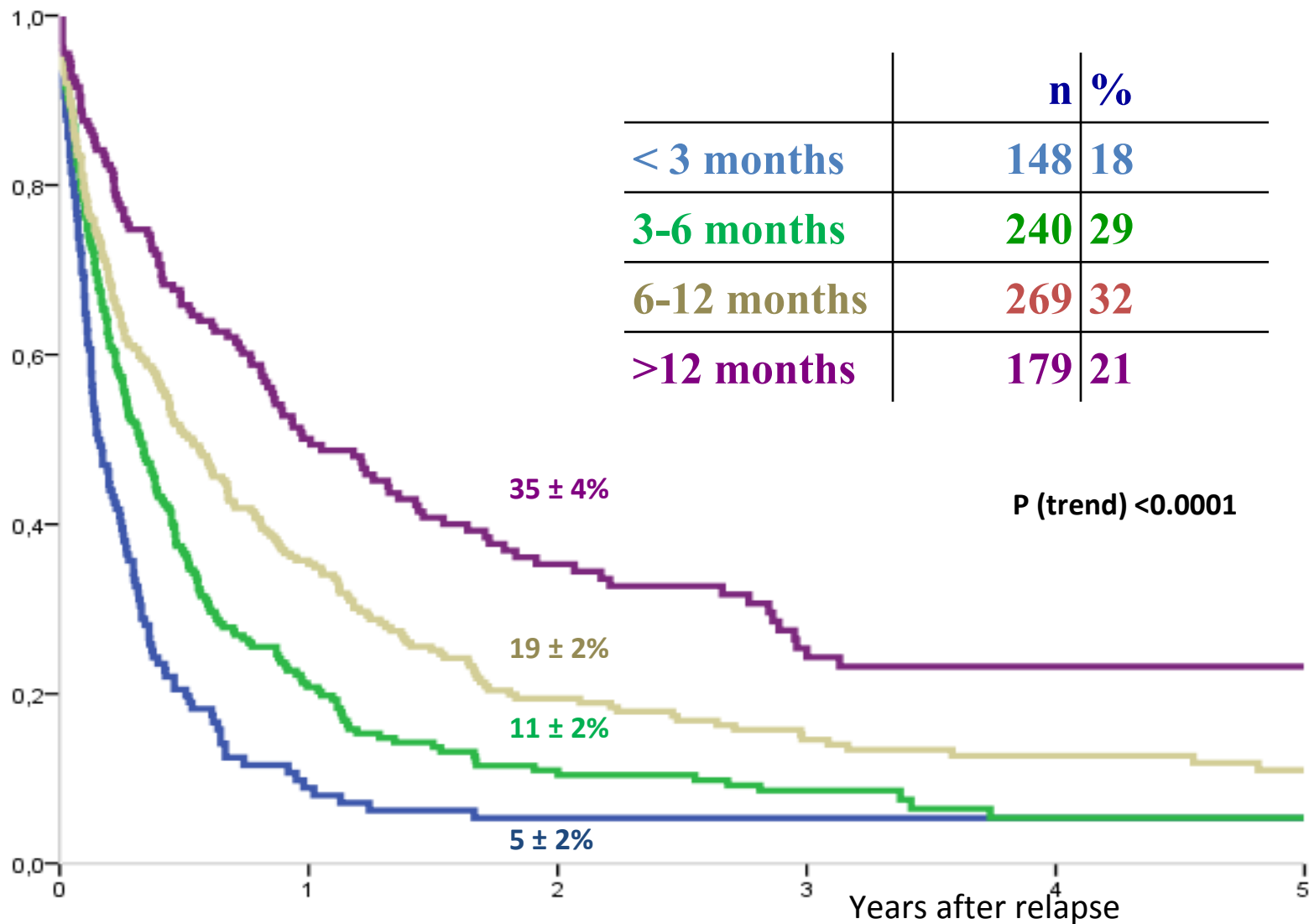
## Overall Survival



§ simulated KM (if the 35 pts – transplanted earlier than 2004, not up-dated after 2005 – all died soon after relapse) OS at 2 years would decrease from 18 to 10%

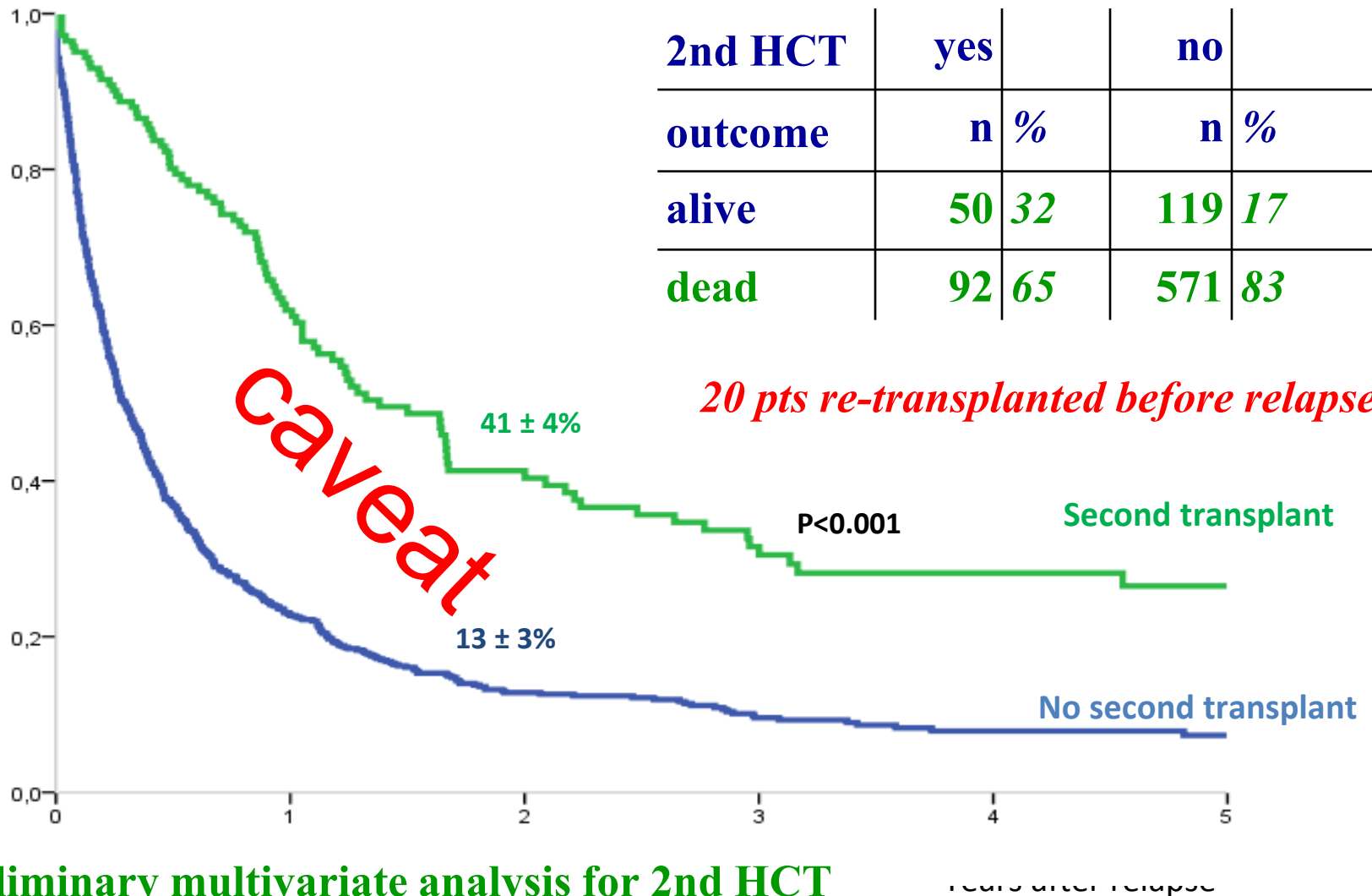
# Relapse after HCT in ALL in pediatric CR1 & CR2 - Results

## Survival according to interval from HCT to relapse



**Relapse after HCT in ALL in pediatric CR1 & CR2 - Results**  
**Outcome after second HCT (n=156)**

| 2nd HCT | yes |    | no  |    |
|---------|-----|----|-----|----|
| outcome | n   | %  | n   | %  |
| alive   | 50  | 32 | 119 | 17 |
| dead    | 92  | 65 | 571 | 83 |



preliminary multivariate analysis for 2nd HCT  
 HR 1.32 (0.98-1.78) p-value 0.072

**Relapse after HCT in pediatric ALL in CR1 & CR2 - Results**  
**Multivariate analysis**

| variable                         |      | HR          | 95% CI      |             | p-value           |
|----------------------------------|------|-------------|-------------|-------------|-------------------|
|                                  |      |             | lower       | upper       |                   |
| <b>second HCT</b>                |      | <b>0.76</b> | <b>0.56</b> | <b>1.02</b> | <b>0.072</b>      |
| <b>interval (ms)</b>             | < 3  | 3.31        | 2.42        | 4.52        | <0.0001           |
| <b>HCT-relapse</b>               | 3-6  | 2.10        | 1.60        | 2.77        | <0.0001           |
| <b>(vs &gt; 12 ms)</b>           | 6-12 | 0.59        | 1.30        | 2.23        | <0.0001           |
| <b>CR2 vs CR1</b>                |      | <b>0.73</b> | <b>1.14</b> | <b>1.67</b> | <b>0.001</b>      |
| <b>T vs B-lineage</b>            |      | <b>1.69</b> | <b>1.33</b> | <b>2.13</b> | <b>&lt;0.0001</b> |
| <b>HLA ident vs other donors</b> |      | <b>0.89</b> | <b>0.93</b> | <b>1.35</b> | <b>0.24</b>       |

# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 1st of 4 approaches

> 1985

Palliative care

Chemotherapy

DLI

Second transplant

- 8 months old
- ALL B-I, MLL germline
- frontline Interfant 06

## INTERFANT PROTOCOL

| Interfant | I-06 (HR only) |     |     |     | I-99 (HR according to I-06 only) |     |     |     |
|-----------|----------------|-----|-----|-----|----------------------------------|-----|-----|-----|
|           | CCR            | Rel | TRM | tot | CCR                              | Rel | TRM | tot |
| MSD       | 2              | 0   | 1   | 3   | 1                                | 4   | 0   | 5   |
| MD        | 12             | 1   | 8   | 21  | 4                                | 2   | 1   | 7   |
| MMD       | 1              | 2   | 2   | 5   | 3                                | 1   | 0   | 4   |
| tot       | 15             | 3   | 11  | 29  | 8                                | 7   | 1   | 16  |



# **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

## **1st of 4 approaches**

- **8 months old (Dec 2012); ALL B-I, MLL germline→ frontline Interfant 06**
- **17 months after diagnosis: relapse ALL B-II (WBC  $60 \times 10^3/\text{mm}^3$ ; Hb 10,3 g/dl; PTL 77000/ $\text{mm}^3$ ; blasts 85%)→ AIEOP REC03**
- **HSCT MUD in 2° CR (5 months after relapse), HLA 10/10; source BM; NCT:  $5 \times 10^8/\text{kg}$ ,  $\text{CD}34^+$   $7 \times 10^6/\text{Kg}$ ; conditioning Bu, Flu, TT; GvHD prophylaxis: CyA, MTX, ATG**
- **aGvHD skin, grade 3, stage II→ mPDN 1mg/Kg (discontinued day +76);**
- **EBV positivity→ 3 Rituximab**

# **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

## **1st of 4 approaches**

- **8 months old (Dec 2012); ALL B-I, MLL germline → frontline Interfant 06**
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- **aGvHD skin, grade 3, stage II → mPDN 1mg/Kg (discontinued day +76);**
- **EBV positivity → 3 Rituximab**
- **MRD positivity 0,8% → IS discontinued day +110 → 3 weeks later: mixed chimerism (PB 10% recipient) and morphological relapse (BM blasts 70%) at 4 months after transplant**
- **palliation, as per parental decision**

# **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

## **1st of 4 approaches: PALLIATION**

**> 1985**

**Palliative care**

**Chemotherapy**

**DLI**

**Second transplant**

**Case 1 - 2014 palliation**

**Case 2 - 2014 ?**



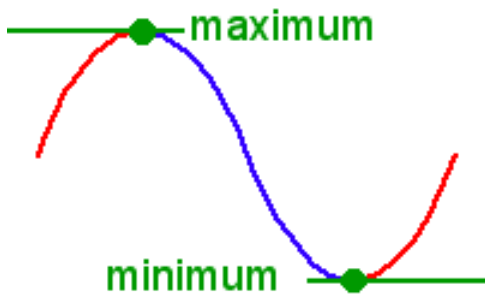
## **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

### **2nd of 4 approaches**

- **LLA common, SNC neg, trasl neg → AIEOP BFM ALL 2009, SER → HR; OT;**
- **+32 ms: isol BM relapse → AIEOP REC 2003.**
- **+5 ms post relapse:**
- **HSCT MSD BM; conditioning TBI, VP16; GvHD prophylaxis: CyA (MTX not given, high risk of relapse)**
- **MRD at transplant  $1.2 \times 10^{-3}$**

# MINIMAL residual disease & transplantation: *explained to high school students...and husband*

- **Philosophy** – Blaise Pascal: “*Infiniment petits*”
- **Math**



The **derivative** of  $f(x)$  at the point  $x$  is equal to the slope of the tangent to  $y = f(x)$  at  $x$

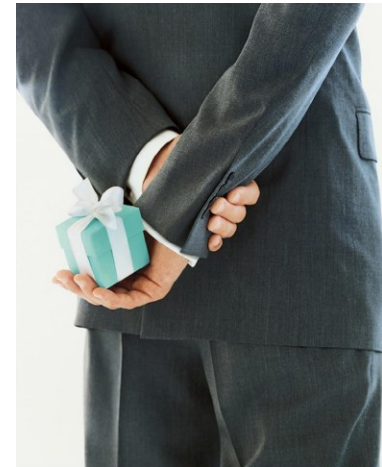
**local minimum** where the graph changes from decreasing to increasing: at this point the tangent has **zero slope**

$$\bullet \lim_{x \rightarrow c} f(x) = L$$

- **Everyday life**

A man decided to buy a diamond:

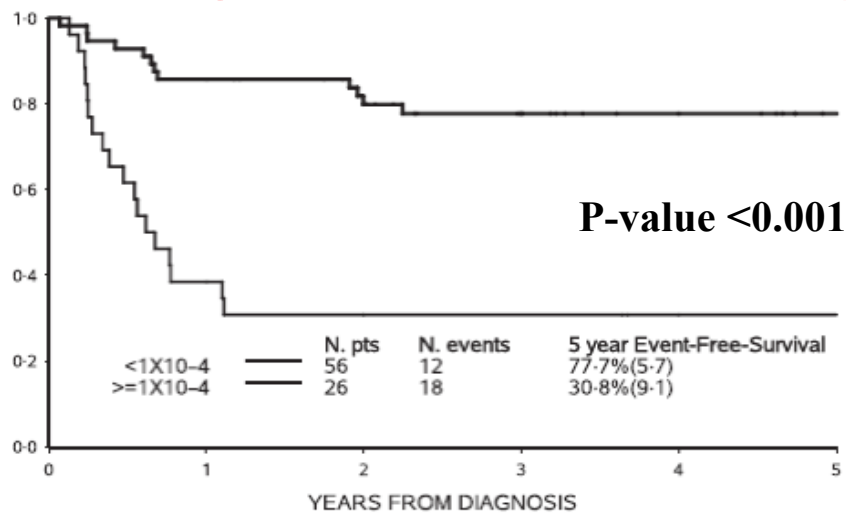
*“I wonder how such a small thing can cost so much...”*



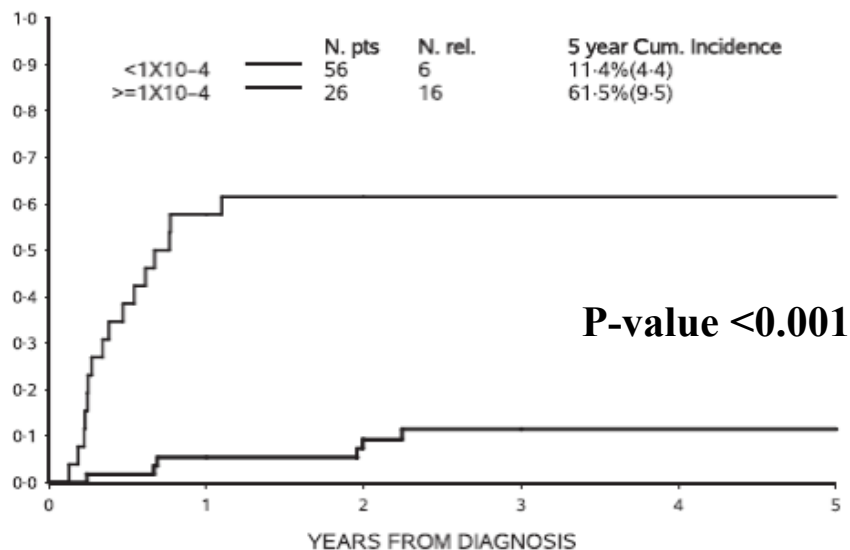
**MRD at transplantation: 56 (68%) neg or  $<1 \times 10^{-4}$  vs 26 (32%)  $\geq 1 \times 10^{-4}$**

**Outcome according to MRD level at transplantation:**

**EFS**



**CI  
Relapse**



**bjh** research paper

**Minimal residual disease before and after transplantation for childhood acute lymphoblastic leukaemia: is there any room for intervention?**

Adriano Baldazzi,<sup>1</sup> Lucia Di Maio,<sup>1</sup> Daniela Silvestri,<sup>1,2</sup> Simona Soglia,<sup>3</sup> Sonia Bonanomi,<sup>4</sup> Annillo Ravelli,<sup>5</sup> Valantino Conter,<sup>6</sup> Andrea Biondi,<sup>1,3</sup> Giovanni Cazzaniga<sup>7</sup> and Maria G. Valsecchi<sup>8</sup>

<sup>1</sup>Clinica Pediatrica, Università degli Studi di Milano-Bicocca, Ospedale San Gerardo, <sup>2</sup>Dipartimento di Scienze della Salute, Centro di Ricerca in Biostatistica per l'Epidemiologia Clinica, Università degli Studi di Milano-Bicocca, and <sup>3</sup>Centro Ricerca Trapianti, Clinica Pediatrica, Università degli Studi di Milano-Bicocca, Monza, Italy

Received 9 July 2013; accepted for publication 18 September 2013

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<sup>†</sup>CG, and M.G.V. share the senior author position.

**Summary**

Eighty-two children and adolescents who underwent allogeneic transplantation for acute lymphoblastic leukaemia in remission (period 2001–2011, median follow-up 4.9 years) had been assessed for minimal residual disease (MRD) by real-time quantitative polymerase chain reaction before and at 1, 3, 6, 9 and 12 months after transplantation. Five-year event-free survival (EFS) and cumulative incidence of relapse were 77.7% [standard error (SE) 5.7] and 11.4% (SE 4.4), respectively, for patients with pre-transplant MRD  $< 1 \times 10^{-4}$  (68%), versus 30.8% (SE 9.1;  $P < 0.001$ ) and 61.5% (SE 9.5;  $P < 0.001$ ), respectively, for those with MRD  $\geq 1 \times 10^{-4}$  (32%). Pre-transplant MRD  $\geq 1 \times 10^{-4}$  was associated with a 9.2-fold risk of relapse [95% confidence interval (CI) 3.54–23.86;  $P < 0.001$ ] compared with patients with MRD  $< 1 \times 10^{-4}$ . Patients who received additional chemotherapy pre-transplant to reduce MRD had a fivefold reduction of risk of failure (hazard ratio 0.19, CI 0.05–0.76,  $P = 0.01$ ). Patients who experienced MRD positivity post-transplant did not necessarily relapse (5-year EFS 40.3%, SE 9.3), but had a 2.5-fold risk of failure (CI 1.05–5.75;  $P = 0.04$ ) if any MRD was detected in the first 100 d, which increased to 7.8-fold (CI 2.2–27.78;  $P = 0.002$ ) if detected after 6 months. Anticipated immunosuppression-tapering according to MRD may have improved outcome, nevertheless all patients with post-transplant MRD  $\geq 1 \times 10^{-3}$  ultimately relapsed, regardless of immunosuppression discontinuation or donor-lymphocyte-infusion. In conclusion, MRD before transplantation had the strongest impact on relapse and MRD positivity after transplantation, mostly if detected early and at low levels, did not necessarily imply relapse. Additional intensified chemotherapy and modulation of immunosuppression may reduce relapse risk and improve ultimate outcome.

**Keywords:** acute lymphoblastic leukaemia, childhood, minimal residual disease, allogeneic transplantation, relapse.

Most children affected with acute lymphoblastic leukaemia (ALL) are potentially cured by conventional chemotherapy as frontline treatment (Conter *et al.* 2010; Schapppe *et al.* 2011; Hunger *et al.* 2012; Pai *et al.* 2012). Even at relapse, approximately one-third of patients can be successfully treated with salvage chemotherapy and radiotherapy (Parker *et al.* 2010; Tallen *et al.* 2010; Freyer *et al.* 2011). Allogeneic haematopoietic stem cell transplantation (HSCT) is considered of benefit for approximately 10% of the patients who are at very-high risk at frontline therapy and for the majority

of patients after relapse (Borgmann *et al.* 2003; Baldazzi *et al.* 2005; Chhansky *et al.* 2012; Zhang *et al.* 2012). Nevertheless, relapse remains the most frequent cause of treatment failure after transplantation (Hildreth *et al.* 2005). During the last decade the prognostic role of minimal residual disease (MRD) has been shown in both frontline and relapse protocols, even when MRD was used to stratify patients and tailor risk-adapted therapy (Eckert *et al.* 2001, 2012; Conter *et al.* 2010; Hunger *et al.* 2012). Since 1998, MRD has also been identified as a relevant prognostic factor in the

## Multivariate analysis MRD at transplantation

### Any event

#### MRD at transplant

$\geq 1 \times 10^{-4}$  vs Negative or  $< 1 \times 10^{-4}$

#### Disease Phase at transplant

CR2 or CR3 vs CR1

#### Acute GVHD

Max Grade II-IV vs 0-I

### Relapse

#### MRD at transplant

$\geq 1 \times 10^{-4}$  vs Negative or  $< 1 \times 10^{-4}$

#### Disease Phase at transplant

CR2 or CR3 vs CR1

#### Acute GVHD

Max Grade II-IV vs 0-I

|  | Hazard ratio | 95% CI            | p-value          |
|--|--------------|-------------------|------------------|
| MRD at transplant<br>$\geq 1 \times 10^{-4}$ vs Negative or $< 1 \times 10^{-4}$ | <b>5.5</b>   | <b>2.58-11.52</b> | <b>&lt;0.001</b> |
| Disease Phase at transplant<br>CR2 or CR3 vs CR1                                 | 2.3          | 0.97-5.35         | 0.06             |
| Acute GVHD<br>Max Grade II-IV vs 0-I   | 0.8          | 0.39-1.70         | 0.59             |
| MRD at transplant<br>$\geq 1 \times 10^{-4}$ vs Negative or $< 1 \times 10^{-4}$ | <b>9.2</b>   | <b>3.54-23.88</b> | <b>&lt;0.001</b> |
| Disease Phase at transplant<br>CR2 or CR3 vs CR1                                 | 2.5          | 0.91-6.80         | 0.07             |
| Acute GVHD<br>Max Grade II-IV vs 0-I   | 0.5          | 0.19-1.24         | 0.13             |

# **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

## **2nd of 4 approaches**

- **LLA common, SNC neg, trasl neg → AIEOP BFM ALL 2009, SER → HR; OT;**
- **+32 ms: isol BM relapse → AIEOP REC 2003.**
- **+5 ms post relapse:**
- **HSCT MSD BM; conditioning TBI, VP16; GvHD prophylaxis: CyA (MTX not given, high risk of relapse)**
- **MRD at transplant  $1.2 \times 10^{-3}$**
- **GvHD skin+liver grade II → mPDN 87 days; stop IS Feb '15**
- **+6 ms post HSCT: MRD pos,  $< 1 \times 10^{-4}$**
- **+6 1/2 ms post HSCT: MRD neg**
- **+7 ms post HSCT: MRD pos,  $< 1 \times 10^{-4}$**
- **+7 1/2 ms post HSCT:  $1.6 \times 10^{-4}$**
- **+9 ms post HSCT  $9.5 \times 10^{-4}$**



## MRD after transplantation-multivariate analyses

|   | Hazard ratio | 95% CI     | p-value |
|---|--------------|------------|---------|
| <b><u>Early post transplant MRD</u></b>         |              |            |         |
| MRD 1-3 ms after HCT – pos vs neg               | 2.5          | 1.05-5.75  | 0.04    |
| Disease Phase at HCT - CR2/CR3 vs CR1           | 2.3          | 0.93-5.73  | 0.07    |
| MRD at HCT - $\geq 10^{-4}$ vs Neg/ $< 10^{-4}$ | 5            | 2.13-11.73 | <0.001  |
| Acute GVHD - II-IV vs 0-I                       | 0.7          | 0.32-1.69  | 0.46    |
| <b><u>Late post transplant MRD</u></b>          |              |            |         |
| MRD 6-12 ms after HCT – pos vs neg              | 7.28         | 2.20-27.28 | 0.002   |
| Disease Phase at HCT - CR2/CR3 vs CR1           | 1.9          | 1.51-7.28  | 0.34    |
| MRD at HCT - $\geq 10^{-4}$ vs Neg/ $< 10^{-4}$ | 3.5          | 1.04-11.50 | 0.04    |
| Acute GVHD - II-IV vs 0-I                       | 3.2          | 0.98-10.48 | 0.06    |

**bjh**  
Minimal residual disease before and after transplantation for childhood acute lymphoblastic leukaemia: is there any room for intervention?

**Abstract**  
Objective: To assess the impact of minimal residual disease (MRD) on survival in childhood acute lymphoblastic leukaemia (ALL) after transplantation. Design: Retrospective analysis of 100 patients who had undergone transplantation for ALL. Results: The impact of MRD on survival was assessed by multivariate analysis. Conclusion: MRD is a strong predictor of survival in childhood ALL after transplantation. The impact of MRD on survival is similar to that of other prognostic factors such as disease phase and cytogenetics. The impact of MRD on survival is similar to that of other prognostic factors such as disease phase and cytogenetics. The impact of MRD on survival is similar to that of other prognostic factors such as disease phase and cytogenetics.

# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 2nd of 4 approaches

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- GvHD skin+liver grade II → mPDN 87 days; stop IS Feb '15
- +6 ms post HSCT: MRD pos  $1.6 \times 10^{-4}$
- +9 ms post HSCT  $9.5 \times 10^{-4}$



**+10 ms after 1st transplant → haplo HSCT (mother, BM)**  
**conditioning: TT, Treo, Flu;**  
**GvHD prophylaxis: CyA, MMF, CY (50 mg/kg day +3, +4)**

**“Cy post”**

# **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

## **2nd of 4 approaches**

### **HAPLOIDENTICAL TRANSPLANTATION**

### **WITH POST TRANSPLANT CYCLOPHOSPHAMIDE**

#### **Post transplantation immunologic cyclophosphamide**

**(50 mg/Kg days +2, +3): —> hypothesis:**

- **selectively depletes alloreactive T-cells**
  - (= creates immunogenic tolerance by specific clonal killing of activated mature T-cells)**
    - **from the donor responsible for GVHD and**
    - **from recipient responsible for rejection**
  - **preserves resting memory T-cells essential for immune reconstitution**
- **no other IS before transplant which would prevent clonal expansion after antigenic stimulation by the graft and killing**

#### **Non-myeloablative conditioning regimen**

- **highly immune-suppressive but moderately myelo-suppressive**
- **immunosuppression should be sufficient to allow donor engraftment**
- **a high dose of donor cells increases the probability of engraftment**
- **immunologic recovery may be faster**
- **initial mixed chimerism protective against GVHD by promoting immune tolerance**

## **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

### **2nd of 4 approaches CY POST HAPLO**

## **“CY post” haplo in pediatrics**

- Yesilipek, Turkey, [Pediater Transplant](#). 2015 Dec 28
- Haploidentical hematopoietic stem cell transplantation with post-transplant high-dose cyclophosphamide in high-risk children: A single-center study
- 15 pts, 16 SCTs
- CY +3, +5; TAC + MMF / PDN
- 6 aGVHD, 2 cGVHD
- 2 TRM
- 12 CCR: OS  $75 \pm 10.8\%$  and DFS  $68.8 \pm 11.6\%$

**... Survey to circulate within EBMT centers**

# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 2nd of 4 approaches: “CY POST” HAPLO

> 1985

Palliative care

Chemotherapy

DLI

Second transplant

Case 1 - 2014  
palliation

Case 2 - 2015  
2nd transplant, haploidentical CY post



# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 3rd of 4 approaches

- ALL B-I → AIEOP R2006;
- + 33 ms: relapse (BM + right testis) → AIEOP LLA REC 2003+orchiectomy; HSCT MD; GvHD gut+liver
- +12 ms after SCT: 2° relapse (left testis + BM MRD positivity)
- 3 ms later: 3° relapse (BM 19% blasts) → chemotherapy (steroid, vincristine, FLAG-D, FLA-G)
- 5 ms later: 4° relapse (BM blasts 41%) → 1 **blinatumomab** → morphological remission (MRD pre-HSCT  $7 \times 10^{-3}$ )
- 3 ms later: haplo (father; BM); conditioning: TT+ Treo+Flu; GvHD prophylaxis: CyA, MMF, CY post
- Nov '15: MRD positivity ( $<1 \times 10^{-4}$ ) → **DLI+CIK**

**Blinatumomab**

## **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

### **3rd of 4 approaches: BLINATUMOMAB**

- **Blinatumomab is a bispecific T-cell engager (BiTE) antibody derived from a B-lineage specific antitumor mouse monoclonal antibody**
- **Blinatumomab is a CD19/CD3 bispecific T-cell engaging (BiTE) antibody that binds to CD3<sup>+</sup> T-cells and co-localizes them with CD19<sup>+</sup> B-cells, thereby activates the T-cells and induces perforin-mediated death of the targeted B-cells**
- **Ongoing randomized COG phase III study for pre-B ALL children in first relapse is scheduled to open in late 2014, combining blinatumomab with UKALLR3 reinduction chemotherapy**

# MRD in HCT for ALL – Results - 7

## Interventions according to MRD

### 4 levels of INTERVENTION:

#### I. CHEMO INTENSIFICATION and HCT postponement

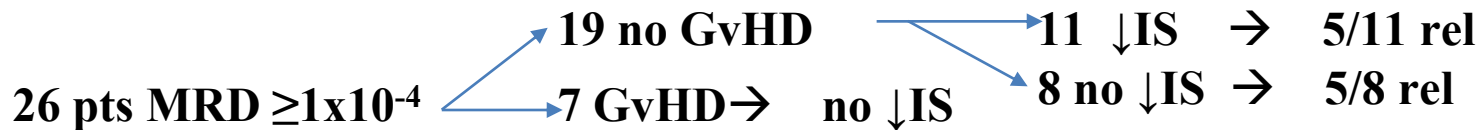
Pre-transplant intervention - based on MRD at the time when HCT was due (pre-pre MRD) (after 2007)



Administration of FLA-D was associated with a **five-fold reduction** of the hazard of **failure** (hazard ratio (HR) 0.19, 95% CI 0.05- 0.70, p-value 0.01)

#### II. EARLY IMMUNOSUPPRESSION TAPERING

Post HCT intervention – based on MRD at transplant (after 2004)





# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 3rd of 4 approaches: blinatumomab

> 1985

Palliative care

Chemotherapy

DLI

Second transplant

2014

palliation

2015

2nd transplant, haploidentical CY post

2015

blinatumomab,

2nd transplant, haploidentical, "CY post"

# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 4th of 4 approaches

- **Thalassemia major** → Sept '09 **HSCT MSD** (donor: HLA-identical sister, source: BM); conditioning: TT-Treo-Flu; GVHD prophylaxis CyA, MTX;
- Decreasing donor chimerism and rejection despite 7 DLI; Dec '09: full autologous reconstitution.
- Apr '11: **Diagnosis of t(9;22) ALL** □ EsPhALL (induction + block HR-1), morphological CR, molecular persistence of disease ( Aug '11: BCR/Abl: 178 copies/10<sup>4</sup> abl copies; MRD <5x10<sup>-4</sup>)
- Sept '11: **2<sup>nd</sup> HSCT MSD** (donor: HLA-identical brother, source BM); conditioning: TT, Treo, Flu; GVHD prophylaxis CyA, MTX; full three-lineage engraftment, full donor chimerism, post-HSCT course uneventful.
- Imatinib since the 2nd month post HSCT, continued due to persistent t(9;22) positivity on PB and BM, despite MRD TCR/Ig negativity.
- Nov '13: relapse as **Bi-phenotypic t(9;22) ALL** in varicella (BM blasts 90%);
- Jan '14: phase I study **nilotinib** enrollment (blasts 50%) □ Feb '14: **CR2**, BM blasts 3%; BCR-ABL 400/10.000 copies;
- May '14: **Stop nilotinib**
- Enrollment in ~~CD19TPALL trial~~ prophylactic arm. CHILDHOPE, P. Amrolia UCL
- Jun '14: **3<sup>rd</sup> HSCT MSD (donor: same HLA-identical brother as for her second transplant, source BM); conditioning: TT, Treo, Flu; GVHD prophylaxis: CyA, MTX);**
- **Post transplant treatment** (Oct- Dec '15): Lymphodepletion with Fludarabin 30mg/m<sup>2</sup>; transfusion of cytotoxic T-cells (CD19-zeta EBV-CTLs); BLCL-Vaccination (Irradiated donor-derived B Lymphoblastoid Cell Line, BLCL);
- **Jan '15: molecular Relapse** □ **May '15 Morphological relapse**

# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 4th of 4 approaches

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- Apr '11: **Diagnosis of t(9;22) ALL** □ EsPhALL (induction + block HR-1), morphological CR, molecular persistence of disease ( Aug '11: BCR/Abl: 178 copies/10<sup>4</sup> abl copies; MRD <5x10<sup>-4</sup>)
- Sept '11: **2<sup>nd</sup> HSCT MSD** (donor: HLA-identical brother, source BM); conditioning: TT, Treo, Flu; GVHD prophylaxis CyA, MTX; full three-lineage engraftment, full donor chimerism, post-HSCT course uneventful.
- **Enrollment in CD19TPALL trial** (relapse in relapse post HSCT, continued due to persistent t(9;22) positivity in PL and BM, despite MRD PCR/Ig negativity).
- Nov '13: relapse as **Bi-phenotypic t(9;22) ALL** in varicella (BM blasts 90%);
- **prophylactic arm: CHILDDHOPE, P. Amrolia UCL**
- Jun '14: Phase I study of imatinib relapse (11 pts 50%) □ Feb '14: CR, BM blasts 3%; ECR, ABL 400/10.000 copies;
- **Stop nilotinib**
- **Enrollment in CD19TPALL trial** prophylactic arm: CHILDDHOPE, P. Amrolia UCL
- Jun '14: **3<sup>rd</sup> HSCT MSD** (donor: same HLA-identical brother as for her second transplant, source BM); conditioning: TT, Treo, Flu; GVHD prophylaxis: CyA, MTX);
- **Post transplant treatment** (Oct- Dec '15): Lymphodepletion with Fludarabin 30mg/m<sup>2</sup>; transfusion of cytotoxic T-cells (CD19-zeta EBV-CTLs); BLCL-Vaccination (Irradiated donor-derived B Lymphoblastoid Cell Line, BLCL);
- **Jan '15: molecular Relapse** □ **May '15 Morphological relapse**

# From hope to reality!

The New York Times

December 9, 2012

## In Girl's Last Hope, Altered Immune Cells Beat Leukemia

By DENISE GRADY

PHILIPSBURG, Pa. — Emma Whitehead has been bounding around the house lately, practicing somersaults and rugby-style tumbles that make her parents wince.

It is hard to believe, but last spring Emma, then 6, was near death from leukemia. She had relapsed twice after chemotherapy, and doctors had run out of options.

The treatment very nearly killed her. But she emerged from it cancer-free, and about seven months later is still in complete remission. She is the first child and one of the first humans ever in whom new techniques have achieved a long-sought goal — giving a patient's own immune system the lasting ability to fight cancer.

# How to cure cancer: a new path...

## Emma Whitehead: the 1st ALL child cured by



THE NEW ENGLAND JOURNAL of MEDICINE

### BRIEF REPORT

## Chimeric Antigen Receptor–Modified T Cells for Acute Lymphoid Leukemia

Stephan A. Grupp, M.D., Ph.D., Michael Kalos, Ph.D., David Barrett, M.D., Ph.D., Richard Aplenc, M.D., Ph.D., David L. Porter, M.D., Susan R. Rheingold, M.D., David T. Teachey, M.D., Anne Chew, Ph.D., Bernd Hauck, Ph.D., J. Fraser Wright, Ph.D., Michael C. Milone, M.D., Ph.D., Bruce L. Levine, Ph.D., and Carl H. June, M.D.

### RESEARCH ARTICLE

#### CANCER IMMUNOTHERAPY

## CD19-Targeted T Cells Rapidly Induce Molecular Remissions in Adults with Chemotherapy-Refractory Acute Lymphoblastic Leukemia

Renier J. Brentjens,<sup>1,2,3,\*</sup> Marco L. Davila,<sup>1†</sup> Isabelle Riviere,<sup>1,2,3,4†</sup> Jae Park,<sup>1</sup> Xiuyan Wang,<sup>3,4</sup> Lindsay G. Cowell,<sup>5</sup> Shirley Bartido,<sup>4</sup> Jolanta Stefanski,<sup>4</sup> Clare Taylor,<sup>4</sup> Malgorzata Olszewska,<sup>4</sup> Oriana Borquez-Ojeda,<sup>4</sup> Jinrong Qu,<sup>4</sup> Teresa Wasielewska,<sup>4</sup> Qing He,<sup>4</sup> Yvette Bernal,<sup>1</sup> Ivelisse V. Rijo,<sup>6</sup> Cyrus Hedvat,<sup>6</sup> Rachel Kobos,<sup>7</sup> Kevin Curran,<sup>7</sup> Peter Steinhilber,<sup>7</sup> Joseph Jurcic,<sup>1</sup> Todd Rosenblatt,<sup>1</sup> Peter Masiak,<sup>1</sup> Mark Frattini,<sup>1</sup> Michel Sadelain<sup>1,2,3,\*</sup>



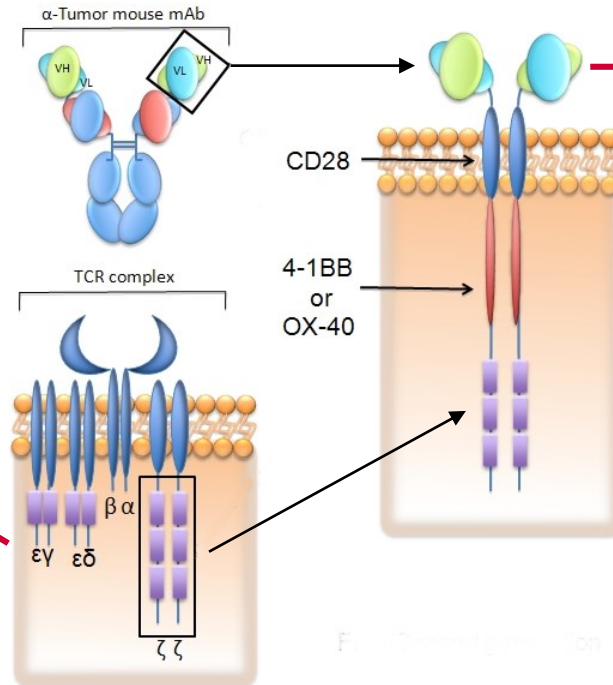
# PEDIATRIC ALL RELAPSING AFTER TRANSPLANT

## 4th of 4 approaches

### Chimeric Receptors for Immunotherapy of Acute Leukemias

#### CHIMERIC ANTIGEN RECEPTORS CARs

An intracellular signaling domain triggering T cell activation



An extracellular domain recognizing tumor-associated antigens derived from mAb



modified from Chekmasova AA, Brentjens RJ (2010), *Discov Med*, 9(44):62-70

# **PEDIATRIC ALL RELAPSING AFTER TRANSPLANT**

## **4th of 4 approaches**

# **CARs T**

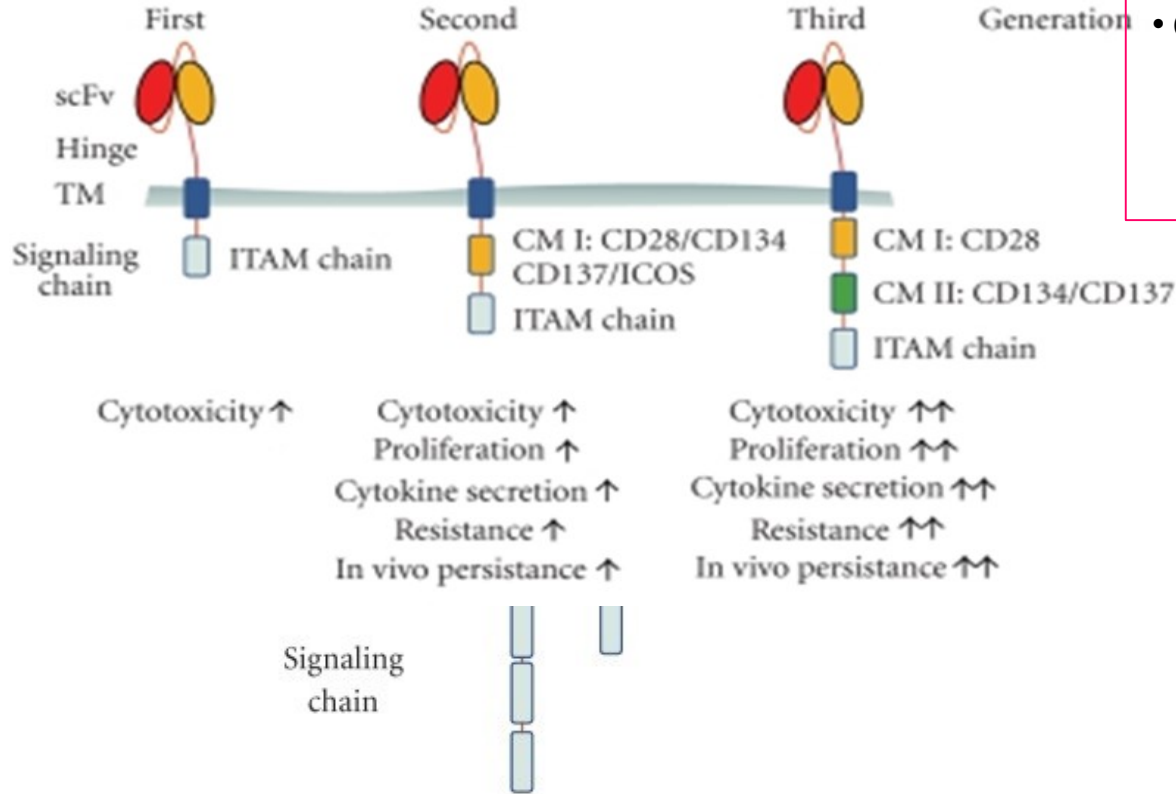
- **Chimeric antigen receptor-modified T-cells (CAR T-cells) with CD19 specificity (adoptive transfer of CD19 $\zeta$  chimaeric receptor transduced donor-derived EBV-specific cytotoxic T-lymphocytes (EBV-CTL)) are generating excitement as a novel therapy for high-risk or relapsed B cell precursor ALL after allogeneic Haematopoietic Stem Cell Transplantation (HSCT).**
- **CAR T cells are patient-derived T-cells, transduced to express a chimeric antigen receptor, which includes an anti-CD19 antibody fragment fused to a T-cell intracellular signaling domain**
- **Second-generation CAR T cells also encode for a costimulatory domain, such as CD28 or members of the tumor necrosis factor receptor family such as CD27, CD137 (4-1BB) and CD134 (OX40). The costimulatory domains activate the CAR T-cells, allowing for targeting and lysis of CD19<sup>+</sup> cells.**

# Chimeric Antigen Receptors (CD19, CD20, CD30, CD33)

**ANTIGEN-BINDING PROPERTY OF ANTIBODIES**

**TCR-TRIGGERING DOMAIN:**

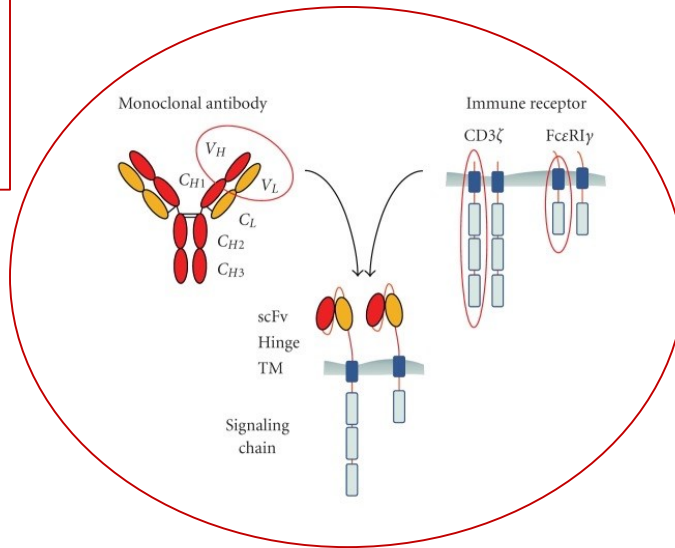
- T-cells activation
- Cytokines secretion
- Cytotoxicity
- Homing





# Redirecting T cell activity with Chimeric Antigen Receptors (CARs)

## ANTIGEN-BINDING PROPERTY OF ANTIBODIES

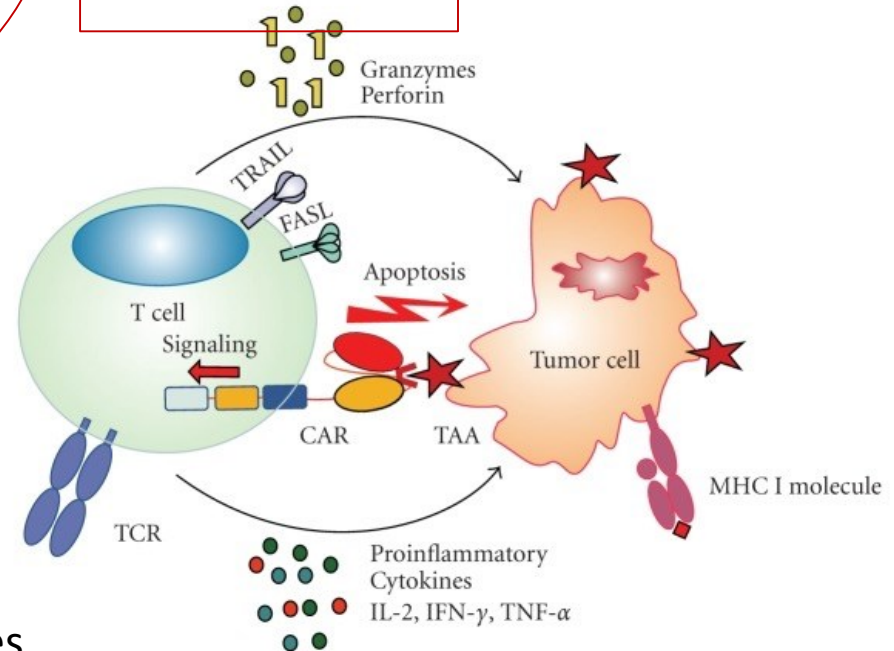


## TCR-TRIGGERING DOMAIN:

- T-cell activation
- Cytokines secretion
- Cytotoxicity
- Persistence
- Homing

## ADVANTAGES OF CARs

- HLA- independent antigen recognition
- Active in both CD4+ and CD8+ T cells
- Target antigens include proteins, carbohydrates and glycolipids
- Immunological memory
- Better biodistribution compared to mAbs



Cartelli et al., 2010

# CD19 CARS AND CLINICAL APPLICATION

## Chemotherapy-Refractory Diffuse Large B-Cell Lymphoma and Indolent B-Cell Malignancies Can Be Effectively Treated With Autologous T Cells Expressing an Anti-CD19 Chimeric Antigen Receptor

James N. Kochenderfer, Mark E. Dudley, Sadik H. Kassim, Robert P.T. Somerville, Robert O. Carpenter, Maryalice Stetler-Stevenson, James C. Yang, Gao Q. Phan, Marybeth S. Hughes, Richard M. Sherry, Mark Raffeld, Steven Feldman, Lily Lu, Yong F. Li, Lien T. Ngo, Andre Goy, Tatyana Feldman, David E. Spaner, Michael L. Wang, Clara C. Chen, Sarah M. Kranick, Avindra Nath, Debbie-Ann N. Nathan, Kathleen E. Morton, Mary Ann Toomey, and Steven A. Rosenberg

## Donor-derived CD19-targeted T cells cause regression of malignancy persisting after allogeneic hematopoietic stem cell transplantation

James N. Kochenderfer,<sup>1</sup> Mark E. Dudley,<sup>2</sup> Robert O. Carpenter,<sup>1</sup> Sadik H. Kassim,<sup>2</sup> Jeremy J. Rose,<sup>1</sup> William G. Telford,<sup>1</sup> Frances T. Hakim,<sup>1</sup> David C. Halverson,<sup>1</sup> Daniel H. Fowler,<sup>1</sup> Nancy M. Hardy,<sup>1</sup> Anthony R. Mato,<sup>3</sup> Dennis D. Hickstein,<sup>1</sup> Juan C. Gea-Banacloche,<sup>1</sup> Steven Z. Pavletic,<sup>1</sup> Claude Sportes,<sup>1</sup> Irina Maric,<sup>4</sup> Steven A. Feldman,<sup>2</sup> Brenna G. Hansen,<sup>1</sup> Jennifer S. Wilder,<sup>5</sup> Bazetta Blacklock-Schuber,<sup>1</sup> Bipulendu Jena,<sup>6</sup> Michael R. Bishop,<sup>7</sup> Ronald E. Gress,<sup>1</sup> and Steven A. Rosenberg<sup>2</sup>

## B-cell depletion and remissions of malignancy along with cytokine-associated toxicity in a clinical trial of anti-CD19 chimeric-antigen-receptor-transduced T cells

James N. Kochenderfer,<sup>1</sup> Mark E. Dudley,<sup>2</sup> Steven A. Feldman,<sup>2</sup> Wyndham H. Wilson,<sup>3</sup> David E. Spaner,<sup>4</sup> Irina Maric,<sup>5</sup> Maryalice Stetler-Stevenson,<sup>6</sup> Gao Q. Phan,<sup>2</sup> Marybeth S. Hughes,<sup>2</sup> Richard M. Sherry,<sup>2</sup> James C. Yang,<sup>2</sup> Udai S. Kammula,<sup>2</sup> Laura Devillier,<sup>2</sup> Robert Carpenter,<sup>1</sup> Debbie-Ann N. Nathan,<sup>2</sup> Richard A. Morgan,<sup>2</sup> Carolyn Laurencot,<sup>2</sup> and Steven A. Rosenberg<sup>2</sup>

## Infusion of donor-derived CD19-redirection virus-specific T cells for B-cell malignancies relapsed after allogeneic stem cell transplant: a phase 1 study

Conrad Russell Y. Cruz,<sup>1</sup> Kenneth P. Micklethwaite,<sup>1</sup> Barbara Savoldo,<sup>1</sup> Carlos A. Ramos,<sup>1</sup> Sharon Lam,<sup>1</sup> Stephanie Ku,<sup>1</sup> Oumar Diouf,<sup>1</sup> Enli Liu,<sup>1</sup> A. John Barrett,<sup>2</sup> Sawa Ito,<sup>2</sup> Elizabeth J. Shpall,<sup>3</sup> Robert A. Krance,<sup>1,4</sup> Rammurti T. Kamble,<sup>1,4</sup> George Carrum,<sup>1,4</sup> Chitra M. Hosing,<sup>3</sup> Adrian P. Gee,<sup>1</sup> Zhuyong Mei,<sup>1</sup> Bambi J. Grilley,<sup>1</sup> Helen E. Heslop,<sup>1,4</sup> Ciona M. Rooney,<sup>1</sup> Malcolm K. Brenner,<sup>1,4</sup> Catherine M. Bollard,<sup>1,4</sup> and Gianpietro Dotti<sup>1</sup>

## Infusing CD19-Directed T Cells to Augment Disease Control in Patients Undergoing Autologous Hematopoietic Stem-Cell Transplantation for Advanced B-Lymphoid Malignancies

Partow Kebriaei,<sup>1</sup> Helen Huls,<sup>2</sup> Bipulendu Jena,<sup>2</sup> Mark Munsell,<sup>1</sup> Rineka Jackson,<sup>2</sup> Dean A. Lee,<sup>2,3</sup> Perry B. Hackett,<sup>4</sup> Gabriela Rondon,<sup>1</sup> Elizabeth Shpall,<sup>1</sup> Richard E. Champlin,<sup>1</sup> and Laurence J.N. Cooper<sup>2,3</sup>

## Efficacy and Toxicity Management of 19-28z CAR T Cell Therapy in B Cell Acute Lymphoblastic Leukemia

Marco L. Davila<sup>1</sup>, Isabelle Riviere<sup>1,2,3,4</sup>, Xiuyan Wang<sup>4</sup>, Shirley Bartido<sup>4</sup>, Jae Park<sup>1</sup>, Kevin Curran<sup>5</sup>, Stephen S. Chung<sup>1</sup>, Jolanta Stefanski<sup>4</sup>, Oriana Borquez-Ojeda<sup>4</sup>, Malgorzata Olszewska<sup>4</sup>, Jinrong Qu<sup>4</sup>, Teresa Wasielewska<sup>4</sup>, Qing He<sup>4</sup>, Mitsu Fink<sup>4</sup>, Himaly Shinglot<sup>4</sup>, Maher Youssif<sup>4</sup>, Mark Satter<sup>4</sup>, Yongzeng Wang<sup>4</sup>, James Hosey<sup>4</sup>, Hilda Quintanilla<sup>1</sup>, Elizabeth Halton<sup>1</sup>, Yvette Bernal<sup>1</sup>, Diana C. G. Bouhassira<sup>2</sup>, Maria E. Arcila<sup>6</sup>, Mithat Gonen<sup>7</sup>, Gail J. Roboz<sup>8</sup>, Peter Maslak<sup>1</sup>, Dan Douer<sup>1</sup>, Mark G. Frattini<sup>9</sup>, Sergio Giralt<sup>1,2</sup>, Michel Sadelain<sup>1,2,3,\*</sup> and Renier Brentjens<sup>1,2,3,\*</sup>

## Safety and persistence of adoptively transferred autologous CD19-targeted T cells in patients with relapsed or chemotherapy refractory B-cell leukemias

\*Renier J. Brentjens,<sup>1,3</sup> Isabelle Rivière,<sup>1,4</sup> Jae H. Park,<sup>1,2</sup> Marco L. Davila,<sup>1,2</sup> Xiuyan Wang,<sup>2,4</sup> Jolanta Stefanski,<sup>2,4</sup> Clare Taylor,<sup>2,4</sup> Raymond Yeh,<sup>1,2</sup> Shirley Bartido,<sup>2,3</sup> Oriana Borquez-Ojeda,<sup>2,4</sup> Malgorzata Olszewska,<sup>2,4</sup> Yvette Bernal,<sup>1</sup> Hollie Pegram,<sup>1,2</sup> Mark Przybylowski,<sup>2,4</sup> Daniel Hollyman,<sup>2,4</sup> Yelena Usachenko,<sup>1,2</sup> Domenick Pirraglia,<sup>2,4</sup> James Hosey,<sup>2,4</sup> Elmer Santos,<sup>3,5</sup> Elizabeth Halton,<sup>1</sup> Peter Maslak,<sup>1</sup> David Scheinberg,<sup>1,3</sup> Joseph Jurcic,<sup>1</sup> Mark Heaney,<sup>1</sup> Glenn Heller,<sup>6</sup> Mark Frattini,<sup>1</sup> and Michel Sadelain<sup>1,3</sup>

## CD19-targeted T cells rapidly induce molecular remissions in adults with chemotherapy-refractory acute lymphoblastic leukemia

Renier Brentjens<sup>\*,†,1,2,3</sup>, Marco L Davila<sup>†,1</sup>, Isabelle Riviere<sup>†,1,2,3,4</sup>, Jae Park<sup>1</sup>, Xiuyan Wang<sup>3,4</sup>, Lindsay G Cowell<sup>7</sup>, Shirley Bartido<sup>4</sup>, Jolanta Stefanski<sup>4</sup>, Clare Taylor<sup>4</sup>, Malgorzata Olszewska<sup>4</sup>, Oriana Borquez-Ojeda<sup>4</sup>, Jinrong Qu<sup>4</sup>, Teresa Wasielewska<sup>4</sup>, Qing He<sup>4</sup>, Yvette Bernal<sup>1</sup>, Ivelise V Rijo<sup>5</sup>, Cyrus Hedvat<sup>5</sup>, Rachel Kobos<sup>6</sup>, Kevin Curran<sup>6</sup>, Peter Steinherz<sup>6</sup>, Joseph Jurcic<sup>1</sup>, Todd Rosenblat<sup>1</sup>, Peter Maslak<sup>1</sup>, Mark Frattini<sup>1</sup>, and Michel Sadelain<sup>\*,1,2,3</sup>

## Chimeric Antigen Receptor-Modified T Cells for Acute Lymphoid Leukemia

Stephan A. Grupp, M.D., Ph.D., Michael Kalos, Ph.D., David Barrett, M.D., Ph.D., Richard Aplenc, M.D., Ph.D., David L. Porter, M.D., Susan R. Rheingold, M.D., David T. Teachey, M.D., Anne Chew, Ph.D., Bernd Hauck, Ph.D., J. Fraser Wright, Ph.D., Michael C. Milone, M.D., Ph.D., Bruce L. Levine, Ph.D., and Carl H. June, M.D.



# PEDIATRIC ALL RELAPSing AFTER TRANSPLANT

## 4 approaches

> 1985

Palliative care

Chemotherapy

DLI

Second transplant

2014

palliation

2015

2nd transplant, haploidentical CY post

2015

blinatumomab,

2nd transplant, haploidentical, "CY post"

2014

2nd transplant, CAR

# NUOVI FARMACI E TRAPIANTO: LLA

What's crucial in pediatric transplantation in ALL

“Standard” innovative transplantation:

- patient selection
- reduce mortality
- reduce long-term sequelae

... and what's next...

**FORUM STUDY**

For omitting Radiation Under Majority Age

Innovative strategies:

Blinatumomab

Post-Cy Haplo

CAR